Stereoselective synthesis of α -O-fucosylated serine, threonine, tyrosine and saccharides

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Stereoselective fucosylation of Boc-Ser/Thr/Tyr-OMe 2–4 and several saccharide alcohols 5–7 by coupling with 2-pyridyl tri-O-benzyl-1-thio- β -L-fucosyl donor 1b under iodomethane activation procedure results in the formation of α -linked fucosylated amino acid methyl ester building blocks 8–10 and α -linked fucosyl saccharides 11–13, respectively.

Introduction

A large number of proteins found in nature are N- and/or O-glycosylated, and the structural diversity of the oligosaccharides and the variety of linkages between sugar and protein moieties is enormous.^{1,2} Glycosylation of a protein is known to influence the properties of the protein in many ways.² Glycosylation provides protection against proteolysis, influences uptake of serum proteins of the liver, effects intra-molecular transport of enzymes to lysosomes, determines human blood groups and regulates leukocyte trafficking to sites of inflammation.^{3,4} Tumour cells show aberrant glycosylation of glycosphingolipids and glycoproteins.⁵ A novel type of Oglycosylation where Xyl-Glc and Xyl-Xyl-Glc have been found attached to serine residues has been reported by several authors investigating the N-terminal Epidermal Growth Factor (EGF).6 More recently, L-fucose has been found a-glycosidically linked to serine or threonine residues within the peptide motif Cys-X-X-Gly-Gly-Thr/Ser-Cys.⁶ Harris et al. have identified the tetrasaccharide NeuAc_a(2 \rightarrow 6)Gal β (1 \rightarrow 4)GlcNAc β (1 \rightarrow 3)Fuc α 1→O-linked to serine 61 of Human Factor IX.⁷ Furthermore, L-fucose is known to be an important residue of branched oligosaccharides in glycoconjugates that bind to selectins.8 These interesting biological properties have led to the synthesis of fucosylated peptides9 and saccharides10 by both enzymic11 and chemical methods¹² by organic chemists in order to facilitate study of the biological functions of peptides which are not glycosylated in nature. Synthetic fucosylated glycopeptides might be potential mimetics of selectin-binding oligosaccharides. Chemical synthesis of building blocks required for glycopeptide^{9,12} and fucosyl saccharide synthesis¹⁰ has been achieved by use of fluoro, chloro, bromo, acetoxy, acetamidate and alkylthio fucosyl donors.^{9,10} Proven success^{13,14} in recent years in the development of the iodomethane activation procedure of 2-pyridylthio glycosides for the stereoselective synthesis of a-linked saccharides has prompted us to look into its applicability for the synthesis of α -fucosylated amino acids and saccharides.

Results and discussion

2,3,4-Tri-*O*-acetyl- α -L-fucopyranosyl bromide¹⁵ was treated with 2-mercaptopyridine and K₂CO₃ in acetone–toluene (3:2) at room temperature for 4 h to obtain 2-pyridyl 2,3,4-tri-*O*acetyl-1-thio- β -L-fucopyranoside **1a** as a crystalline compound in 79% yield, mp 120–122 °C; $[a]_{\rm D}$ = 36.0 (*c* 1.0, CHCl₃).† Formation of compound **1a** was evident from the ¹H NMR spectrum by the appearance of the anomeric proton signal at δ 5.8 (d, $J_{1,2}$ 10 Hz) and aromatic protons (4 H) between δ 6.90–8.50. Compound 1a on further reaction with a catalytic amount of sodium methoxide in methanol for 6 h at room temp. resulted in the formation of 2-pyridyl 1-thio- β -L-fucopyranoside (I; R = H) which was allowed to react with NaH-benzyl bromide (BnBr)dimethylformamide (DMF) at 0 °C for 3 h to obtain 2-pyridyl 2,3,4-tri-O-benzyl-1-thio-β-L-fucopyranoside 1b as a crystalline solid, mp 134-136 °C, in 76% yield. Fucosylation of N-Bocserine methyl ester 2[‡] was carried out by reaction with compound 1b in the presence of iodomethane as activator in dichloromethane at 50 °C for 62 h to afford, in high stereocontrol, the a-linked N-Boc-3-O-(2,3,4-tri-O-benzyl-a-L-fucopyranosyl)-L-serine methyl ester 8 in 79% yield, as a syrup, $[a]_{D}$ -31 (c 1.0, CHCl₃). The presence of an α -glycosidic linkage in compound 8 was confirmed by the small coupling constant $J_{1'2'}$ 3.8 Hz in the ¹H NMR spectrum and from the appearance of C-1' at $\delta_{\rm C}$ 94.53 in the ¹³C NMR spectrum. Likewise, N-Bocthreonine methyl ester 3 and N-Boc-tyrosine methyl ester 4 were treated with compound 1b to obtain N-Boc-3-O-(2,3,4tri-O-benzyl-a-L-fucopyranosyl)-L-threonine methyl ester 9 and N-Boc-3-O-(2,3,4-tri-O-benzyl-α-L-fucopranosyl)-L-tyrosine methyl ester 10, respectively, in good yields (73-76%) and which were characterised by their respective ¹H and ¹³C NMR spectra.

Coupling of the glycosyl acceptors 1,2:3,4-di-O-isopropylidene-α-D-galactopyranoside 5, 1,2:5,6-di-O-isopropylidene-α-D-glucofuranose 6 and benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy- α -D-glucopyranose 7 with the 2-pyridylthio donor 1b in dichloromethane containing iodomethane at 50 °C for 48-64 h gave their respective α-linked disaccharides 1,2:3,4-di-Oisopropylidene-6-O-(2,3,4-tri-O-benzyl-α-L-fucopyranosyl)-α-D-galactopyranose 11 as a crystalline solid, mp 115-116 °C $(lit., {}^{10} 116-117 °C), [a]_{D} -118 (c 2.0, CH_2Cl_2) {lit., {}^{10} [a]_{D} -117$ (c 2.0, CH₂Cl₂)}, 1,2:5,6-di-O-isopropylidene-3-O-(2,3,4-tri-Obenzyl-a-L-fucopyranosyl)-a-D-glucofuranose 12 as a syrup, characterised by comparison of the spectral data with those reported in the literature,¹⁰ and benzyl 2-acetamido-4,6-O-benzylidene-2-deoxy-3-O-(2,3,4-tri-O-benzyl-α-L-fucopyranosyl)-a-D-glucopyranoside 13 as a crystalline compound (mp 183 °C), characterised from the ¹H NMR spectral data: appearance of 1'-H signal at δ 5.21 as a doublet with a small coupling $J_{1',2'}$ of 3.5 Hz, and from the ¹³C NMR spectrum: appearance of C-1,1' signals at $\delta_{\rm C}$ 97.2 and 99.4.

In conclusion, yet another highly stereoselective α -fucosylation of amino acid esters and several sugar alcohols has been achieved by iodomethane activation of perbenzylated 2pyridyl thio-L-fucosyl donors.

[†] Units for $[a]_{D}$ -values are $10^{-1} \text{ deg cm}^2 \text{ g}^{-1}$.

[‡] Boc = *tert*-butoxycarbonyl.



Experimental

¹H NMR spectra were measured with a Varian Gemini (200 and 400 MHz) spectrometer, with tetramethylsilane as internal standard for solutions in deuteriochloroform. *J*-Values are given in Hz. ¹³C NMR spectra were taken with a Varian Gemini (50 MHz) spectrometer with CDCl₃ as internal standard ($\delta_{\rm C}$ 77.0) for solutions in deuteriochloroform. Optical rotations were measured with a JASCO DIP-370 instrument. Organic solutions were dried over anhydrous Na₂SO₄ and concentrated below 40 °C *in vacuo.*

2-Pyridyl 2,3,4-tri-O-acetyl-1-thio-β-L-fucopyranoside 1a

To a solution of 2-mercaptopyridine (0.6 g, 5.4 mmol) in dry acetone (15 cm³) was added anhydrous K_2CO_3 (0.75 g, 5.4 mmol) and the mixture was stirred at room temperature for 30 min. α -Acetobromofucose¹⁵ (1.6 g, 4.5 mmol) as a solution in dry toluene (10 cm³) was added to the above reaction mixture and the mixture was stirred for 4 h at room temp. The reaction mixture was diluted with toluene (20 cm³) and filtered, and the

organic phase was washed successively with water and saturated aq. NaHCO₃, dried (anhydrous Na₂SO₄), and concentrated to a residue, which was recrystallised from dichloromethane–hexane to obtain the title compound **1a** (1.3 g, 79%) as needles, mp 120–122 °C (Found: C, 52.32; H, 5.48. C₁₇H₂₁NO₇S requires C, 53.26; H, 5.52%); $[a]_D$ –36.0 (*c* 1.0, CHCl₃); δ_H (200 MHz; CDCl₃) 1.20 (3 H, d, *J* 6.3, 6-H₃), 2.0, 2.1 and 2.2 (9 H, 3 s, OAc), 3.95 (1 H, m, 5-H), 5.10–5.40 (3 H, m, 2-, 3- and 4-H), 5.80 (1 H, d, *J* 10, 1-H) and 6.90–8.50 (4 H, m, ArH).

2-Pyridyl 2,3,4-tri-*O*-benzyl-1-thio-β-L-fucopyranoside 1b

Compound 1a (1.3 g, 3.3 mmol) was deacetylated by reaction with cat. NaOMe-MeOH at room temp. for 6 h, the product was neutralised with carbon dioxide, and the solvent was removed to obtain 2-pyridyl 1-thio-α-L-fucopyranoside (0.8 g, 92.6%), which was dried at 60 °C for 3 h, dissolved in DMF (3 cm³), and sodium hydride (0.33 g, 14 mmol) was added followed by benzyl bromide (1.36 cm³, 11.2 mmol) at 0 °C and the mixture was stirred for 3 h. When TLC indicated completion of the reaction, excess of NaH was decomposed by addition of methanol (1 cm³), and the mixture was diluted with water (100 cm^3) and extracted into dichloromethane (2 × 50 cm^3). The organic phase was washed with water, dried, and concentrated to yield a syrup, which was purified by filtration on a bed of silica gel [60-120 mesh; hexane-ethyl acetate (7:1)] to obtain the title compound 1b (1.2 g, 76%) as yellow needles, mp 134-136 °C (Found: C, 72.79; H, 6.21. C₃₂H₃₃NO₄S requires C, 72.84; H, 6.30%); $[a]_{D}$ – 15 (c 1.0, CHCl₃); δ_{H} (200 MHz; CDCl₃) 1.25 (3 H, d, J 6.3, 6-H₃), 3.55-5.15 (10 H, m, 2-5-H, $3 \times OCH_2Ph$), 5.39 (1 H, d, J 10.0, 1-H) and 6.90–8.50 (19 H, m, ArH).

N-Boc-3-*O*-(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-L-serine methyl ester 8

A mixture of compound 1b (0.2 g, 0.38 mmol), N-(tertbutoxycarbonyl)-L-serine methyl ester 2 (0.08 g, 0.38 mmol) and powdered 4 Å molecular sieves (20 mg) in dry dichloromethane (5 cm³) containing 5% iodomethane was heated to 50 °C for 62 h. Reaction was monitored by TLC and when complete the reaction mixture was filtered on Celite, the filter was washed with ethyl acetate (15 cm³) and the combined filtrate and washings were concentrated to obtain a residue, which was chromatographed [SiO₂; hexane-ethyl acetate (9:1)] to obtain the *title* compound 8 (0.19 g, 79%) as a syrup (Found: C, 67.92; H, 7.04. $C_{36}H_{45}NO_9$ requires C, 68.01; H, 7.14%); $[a]_D -31$ (c 1.0, CHCl₃); $\delta_{\rm H}(400 \text{ MHz}; \text{CDCl}_3)$ 1.05 (3 H, d, J 6.4, 6'-H₃), 1.45 (9 H, s, CMe₃), 3.45-4.80 (12 H, m, 2-H, 3-H₂, 2'-5'-H and 2.5 × OCH₂Ph), 3.69 (3 H, s, OCH₃), 4.82 (1 H, d, J_{1',2'} 3.8, 1'-H), 4.87 (1 H, d, J_{gem} 11.6, OCH₂Ph), 5.81 (1 H, d, J 9.0, NH) and 7.20-7.45 (15 H, m, ArH); $\delta_{\rm C}(50~{\rm MHz};~{\rm CDCl_3})$ 16.52 (C-6'), 28.35 (CMe₃), 52.14 (OCH₃), 54.95 (C-2), 68.90 (C-3), 72.30, 72.98, 73.80 and 74.82 (C-5', 3 × OCH₂Ph), 76.02 (C-2'), 77.45 (C-3'), 79.10 (C-4'), 94.53 (C-1'), 127.3-138.52 (aromatic), 156.34 (C=O, carbamate) and 171.85 (C=O, ester).

N-Boc-3-*O*-(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-L-threonine methyl ester 9

A mixture of compound **1b** (0.3 g, 0.57 mmol), *N*-(tertbutoxycarbonyl)-L-threonine methyl ester **3** (0.12 g, 0.57 mmol) and powdered 4 Å molecular sieves (15 mg) in dry dichloromethane (10 cm³) containing 5% iodomethane was heated to 50 °C for 60 h. When TLC indicated completion of the reaction, the mixture was filtered on Celite, the filter was washed with ethyl acetate (15 cm³) and the combined filtrate and washings were concentrated to obtain a residue, which was chromatographed [SiO₂; hexane–ethyl acetate (9:1)] to obtain the *title compound* **9** (0.18 g, 73%) as a syrup (Found: C, 68.29; H, 7.17. C₃₇H₄₇NO₉ requires C, 68.39; H, 7.29%); $[a]_D -42$ (*c* 1.0, CHCl₃); δ_H (400 MHz; CDCl₃) 1.05 (3 H, d, *J* 6.3, 6'-H₃), 1.19 (3 H, d, *J* 6.7, 4-CH₃), 1.46 (9 H, s, CMe₃), 3.40–4.80 (11 H, m, 2and 3-H, 2'-5'-H and 2.5 × OCH₂Ph), 3.19 (3 H, s, OCH₃), 4.81 (1 H, d, $J_{1',2'}$ 3.8, 1'-H), 4.85 (1 H, d, J_{gem} 12.0, OCH₂Ph), 5.35 (1 H, d, J 9.6, NH) and 7.2–7.45 (15 H, m, ArH); $\delta_{\rm C}$ (50 MHz; CDCl₃) 15.83 (C-4), 16.58 (C-6'), 28.32 (CMe₃), 52.11 (OCH₃), 58.32 (C-2), 66.70 (C-3), 72.11 (C-5'), 74.76, 73.20, 72.97 (3 × OCH₂Ph), 76.0 (C-2'), 77.43 (C-3'), 78.90 (C-4'), 94.84 (C-1'), 127.4–138.52 (arom.), 156.36 (C=O, carbamate) and 171.54 (C=O, ester).

N-Boc-3-O-(2,3,4-tri-O-benzyl- α -L-fucopyranosyl)-L-tyrosine methyl ester 10

A mixture of compound 1b (0.2 g, 0.38 mmol), N-(tertbutoxycarbonyl)-L-tyrosine methyl ester 4 (0.1 g, 0.38 mmol) and powdered 4 Å molecular sieves (15 mg) in dry dichloromethane (5 cm³) containing 5% iodomethane was heated to 50 °C for 78 h. When TLC indicated completion of the reaction, the mixture was filtered on Celite, the filter was washed with ethyl acetate (15 cm³), the combined filtrate and washings were concentrated to obtain a residue, which was chromatographed [SiO₂; hexane-ethyl acetate (9:1)] to obtain the *title compound* **10** (0.2 g, 76%) as a crystalline solid, mp 106–107 °C (Found: C, 71.02; H, 6.77. C42H49NO9 requires C, 71.16; H, 6.82%); [a]D -44 (c 1.0, CHCl₃); $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.05 (3 H, d, J 6.7, 6'-H₃), 1.41 (9 H, s, CMe₃), 3.02 (2 H, m, 3-H₂), 3.71 (3 H, s, OCH₃), 3.94–5.01 (12 H, m, NH, 2-H, 2'-5'-H and $3 \times$ OCH₂Ph), 5.39 (1 H, d, J 3.6, 1'-H) and 6.97-7.41 (19 H, m, ArH); δ_c(50 MHz; CDCl₃) 16.54 (C-6'), 28.23 (CMe₃), 37.49 (C-3), 52.13 (OMe), 67.16 (C-2), 73.26, 74.90, 76.08, 77.62 and 79.18 (C-2'-5', 3 × OCH₂Ph), 96.40 (C-1'), 116.92 (C-6, -8), 127.4-138.8 (aromatic), 156.36 (C=O, carbamate) and 172.30 (C=O, ester).

1,2:3,4-Di-*O*-isopropylidene-6-*O*-(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-α-D-galactopyranose 11

A mixture of compound 1b (0.3 g, 0.57 mmol), 1,2:3,4-di-Oisopropylidene- α -D-galactopyranose 5 (0.15 g, 0.57 mmol) and powdered 4 Å molecular sieves (20 mg) in dry dichloromethane (15 cm³) containing 5% iodomethane was heated to 50 °C for 48 h. Reaction was monitored by TLC and when complete the reaction mixture was filtered on Celite, the filter was washed with ethyl acetate (15 cm³), and the combined filtrate and washings were concentrated to obtain a residue, which was chromatographed [SiO₂; hexane-ethyl acetate (4:1)] to obtain the title disaccharide 11 (0.3 g, 77%) as a crystalline solid, mp 115-116 °C (lit., ¹⁰ 116–117 °C) (Found: C, 69.08; H, 7.05. $C_{39}H_{48}O_{10}$ requires C, 69.21; H, 7.15%); [a]_D -118 (c 2.0, CH₂Cl₂) [lit., $-117 (c 2.0, CH_2Cl_2)]; \delta_H(400 \text{ MHz}; CDCl_3) 1.15 (3 \text{ H}, d, J 6.5,$ 6'-H₃), 1.29, 1.32, 1.42 and 1.5 (12 H, 4 s, 2 × OCMe₂), 3.5–5.0 (17 H, m, 2–5-H, 6-H₂, 1'–5'-H and 3 × OCH₂Ph), 5.51 (1 H, d, J 5.0, 1-H) and 7.2–7.45 (15 H, m, ArH); δ_{c} (50 MHz; CDCl₃) 16.6 (C-6'), 24.4, 24.8, 25.9 and 26.0 (4 × CH₃), 65.2, 66.3, 70.4, 70.7, 72.7, 73.1, 74.6, 76.2, 76.3 and 79.2 (C-2–6, C-2'–5', 3 × OCH₂Ph), 96.1 (C-1'), 97.2 (C-1), 108.3 and 108.9 (2 × OCMe₂) and 127.0-138.0 (aromatic).

1,2:5,6-Di-*O*-iospropylidene-3-*O*-(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-α-D-glucofuranose 12

A mixture of compound **1b** (0.25 g, 0.47 mmol), 1,2:5,6-di-*O*isopropylidene- α -D-glucofuranose **6** (0.13 g, 0.47 mmol) and powdered 4 Å molecular sieves (20 mg) in dry dichloromethane (15 cm³) containing 5% iodomethane was heated to 50 °C for 48 h. Reaction was monitored by TLC and when complete the reaction mixture was filtered on Celite, the filter was washed with ethyl acetate (10 cm³), and the combined filtrate and washings were concentrated to obtain a residue, which was chromatographed [SiO₂; hexane–ethyl acetate (5:1)] to obtain the title disaccharide **12** (0.27 g, 85%) as a syrup (Found: C, 69.12; H, 7.04. Calc. for C₃₉H₄₈O₁₀: C, 69.21; H, 7.15%); [*a*]_D –102 (*c* 2.0, CHCl₃) [lit.,¹⁰ –98 (*c* 2.0, CHCl₃)]; $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.10 (3 H, d, *J* 6.0, 6'-H₃), 1.22, 1.25, 1.36 and 1.46 (12 H, 4 s, $2 \times OCMe_2$), 3.5–5.0 (17 H, m, 2–5-H, 6-H₂, 1'–5'-H and $3 \times OCH_2$ Ph), 5.80 (1 H, d, *J* 4.6, 1-H) and 7.2–7.45 (15 H, m, ArH); δ_C (50 MHz; CDCl₃) 16.8 (C-6'), 25.6, 26.5, 26.8 and 27.0 (4 × CH₃), 67.2, 68.1, 72.4, 73.3, 74.0, 75.1, 76.7, 78.1, 79.5, 81.4 and 82.5 (C-2–6, C-2'–5', 3 × OCH₂Ph), 96.2 (C-1'), 105.6 (C-1), 109.3 and 112.1 (2 × OCMe₂) and 127–139 (aromatic).

Benzyl 2-acetamido-4,6-*O*-benzylidene-2-deoxy-3-*O*-(2,3,4-tri-*O*-benzyl-α-L-fucopyranosyl)-α-D-glucopyranoside 13

A mixture of compound 1b (0.155 g, 0.29 mmol), benzyl 2acetamido-4,6-O-benzylidene-2-deoxy- α -D-glucopyranoside 7 (0.118 g, 0.29 mmol) and powdered 4 Å molecular sieves (10 mg) in dry dichloromethane (10 cm³) containing 5% iodomethane was heated to 50 °C for 60 h. Reaction was monitored by TLC and when complete the reaction mixture was filtered on Celite, the filter was washed with ethyl acetate (10 cm³), and the combined washings and filtrate concentrated to obtain a residue, which was chromatographed [SiO₂; hexane-ethyl acetate (3:1)] to obtain the title disaccharide 13 (0.18 g, 76%) as a crystalline solid, mp 183-184 °C (Found: C, 73.45; H, 6.62. C49H53NO9 requires C, 73.56; H, 6.67%); $[a]_{\rm D}$ -30.6 (c 1.0, CHCl₃); $\delta_{\rm H}$ (400 MHz; CDCl₃) 1.07 (3 H, d, J 3.67, 6'-H₃), 1.25 [3 H, br s, N=C(OH)CH₃], 3.62-5.0 (18 H, m, 2-5-H, 6-H₂, 2'-5'-H and 4 × OCH₂Ph), 5.19 and 5.21 (2 H, 2 d, J 3.12 and 3.57, 1- and 1'-H), 5.59 (1 H, s, O₂CHPh) and 7.2-7.45 (25 H, m, ArH); δ_c(50 MHz; CDCl₃) 16.7 (C-6'), 22.4 [N=C(OH)CH₃], 54.0, 63.2, 67.6, 68.8, 70.0, 73.1, 74.5, 74.9, 75.8, 77.5, 78.1 and 82.0 (C-2-6, C-2'-5' and $4 \times OCH_2Ph$), 97.2 and 99.4 (C-1, -1'), 100.0 (O₂*C*HPh), 125–138 (aromatic) and 170.3 (C=O).

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